Welcome to Plant and Microbial Sciences

# **Course Aims and Philosophy**

 Provide an integrated overview of plant and microbial biology

 Address all levels from molecules to ecological communities





## Lecture Content Overview

- Fundamental aspects of plant biology and microbiology
- Related to current world issues e.g.
  - Biofuels
  - Crop protection
  - Climate change





# Practical Classes I

- Brand new teaching lab
- Integrated practicals:
  - Make your own GM plant containing a reporter gene
  - Physiology of tobacco with Rubsico antisense constructs
  - Plant Pathology





# Practical Classes II

Visits to:

- Botanical Garden
- NIAB Innovation Farm
- Local Field Sites (e.g. Hayley Wood)



# Portugal Field Trip

- Mini projects
- See lecture material out in the field
- Sunshine!
- 15th March-22nd March 2020
- Sign up on Moodle





# Support on Moodle

- Lecture and practical material
- Glossaries for every lecture block
- Interactive resources for consolidation

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#### Welcome to the NST IB Plant and Microbial Sciences 2018-19



This site provides learning resources for students taking IB Plant and Microbial Sciences including; Lecture timetable, Course information, Supplementary lecture material, Student feedback and Course management.

If you have any questions about any of this content, or want to suggest additional ideas for content please email (ugadmin@plantsci.cam.ac.uk)

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🕂 💿 Electronic Timetable 💉	Edit 👻
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## NST PMS 1B: Origins of modern agriculture

Prof. Jim Haseloff (jh295): Supplementary lecture materials at haseloff.plantsci.cam.ac.uk

#### Lecture 1. Plant breeding and transformation

- (i) Crop domestication, with maize as an example
- (ii) Modern agriculture, hybrid maize and the rise of agribusiness
- (iii) Green Revolution
- (iv) Agrobacterium mediated plant transformation

#### Lecture 2. From genotype to phenotype

- (i) Designing synthetic plant genes
- (ii) Single gene traits: pest and herbicide resistance
- (iii) Reporter genes
- (iv) Microscopy

#### Lecture 3. Crop traits

- (i) Complex traits and breeding
- (ii) Cellular growth
- (iii) Trait development in Brassicas
- (iv) Pod shatter in Arabidopsis and Brassica crops.

Following lectures: CO<sub>2</sub> levels, photosynthesis and carbon capture (Hibberd); Nutrient availability (Davies); Global warming: Drought and water relations (Griffiths); Temperature responses

## NST PMS 1B

## Lecture 1: Plant breeding and transformation

### **Origins of world crops**





#### Human migration and establishment of population centres

**Recreation of an Aztec market as seen by first Europeans** 





### **Domestication of maize**



Figure 1. Domestication of corn. The upper image shows the mature inflorescence, or "ear" of teosinte (*Zea mays* ssp. *mexicana*), the probable wild progenitor of modern corn (or maize, *Zea mays* ssp. *mays* L.), shown in the lower image. The teosinte inflorescence has no cob, allowing the seed to separate and disperse easily when they are mature. Selection over time by early agriculturalists resulted in types that retained their seed on the ear, leading to the development of the cob. Modern breeding has greatly increased the size and number of seed per ear. (Courtesy J. Doebley, University of Wisconsin)







### **Maize breeding**



 Natural variation occurs in the wild population.



 Seeds for the next generation are chosen only from individuals with the most desirable traits.



Repeat this process for several generations.



Over time, the quality of the crop increases.

Sculpture of Mexica Goddess Chicomecoatl with Ears of Corn Museum of Anthropology - Mexico City - Mexico

### Examples of some of the 59 native Mexican maize landraces.





## Maize diversity spread across South and North America



Maize domestication was accompanied by modification of many plant traits related to agronomy, growth and yield



#### Major differences between maize and teosinte map to few loci

Table 1.	List of principal tra	aits distinguishing maize and teosinte	

Trait	Description
CUPR (cupules per rank)	Number of cupules in a single rank
DISA (disarticulation score)	Tendency of ear to shatter (1-10 scale)
GLUM (glume score)	Hardness and angle of outer glume (1-10 scale)
LBIL (lateral branch internode)	Average length of internodes on the primary lateral branch
LIBN (branch number)	Number of branches in primary lateral inflorescence
PEDS (pedicellate spikelet score)	Percentage of cupules lacking the pedicellate spikelet
PROL (prolificacy)	Number of ears on the primary lateral branch
RANK (rank)	Number of rows of cupules
STAM (staminate score)	Percentage of male spikelets in primary lateral inflorescence



#### **Genetic crossing to produce hybrid Maize**

FIRST YEAR



FIGURE 4.—Diagram of method of crossing inbred plants and the resulting single crosses to produce double-cross hybrid seed. A field grown from such hybrid seed is shown on the cover of this bulletin.



Roswell Garst: marketing and adoption of hybrid maize.

Growth of seed companies (like Garst Seed) and increasing use of fertilisers and pesticides.

Beginning of modern agriculture and integration of industrialised approaches to food production.





## Maize is the world's most successful crop



Average regional maize output (kg/ha)

Norman Borlaug and the Green Revolution





## **The Green Revolution**





CHART 4: Africa missed out on the Green Revolution.

Primary Gene Pool Same and closely related species. Complete chromosome pairing. Crosses by pollination successful. Secondary Gene Pool More-distant species. Incomplete chromosome pairing. Partial F1 sterility. Crosses by pollination difficult; may require embryo culture. 1° Tertiary Gene Pool Marginally sexually compatible species. Severe F1 sterility. Crosses generally not successful. Quaternary Gene Pool All organisms, including animals and microbes. Crosses not possible by pollination or tissue culture methods. Gene transfer via transgenic methods.

#### **Expansion of the gene pool**

Figure 4. Sources of genetic variation for crop improvement. Breeders produce plants with improved combinations of genes by crossing (hybridization) and selection within the primary gene pool, which is comprised of a crop species and its closest related wild species. Tissue culture methods such as embryo culture are commonly used to enable genes from the secondary gene pool to be transferred into the cultivated species. Other methods such as somatic hybridization sometimes allow genes from the tertiary gene pool of more distantly related species to be transferred into crop plants. The immense gene resources of the quaternary gene pool (essentially all other organisms) can be used for crop improvement only via transgenic methods.



## **Crown gall disease**



#### Agrobacteriun tumefaciens is the causal agent of crown gall disease





Agrobacterium transfers genes for tumour growth and opine biosynthesis to plant cells

Agrobacterium tumefaciens exploits a modified bacterial conjugation system in order to transform susceptible plant cells.



T4SS = Type IV secretion system



#### Fig. 1.

Schematic of octopine-type Ti plasmid pTiA6 showing locations of genes coding for plasmid maintenance (*rep*), infection of plant cells (*vir* region, T-DNA), cell survival in the tumor environment (opine catabolism), and conjugative transfer of the Ti plasmid to recipient agrobacteria (*tra* and *trb*). The various contributions of the *vir* gene products to T-



#### **Fig. 4.**

Genetic organization of the *A. tumefaciens* Ti plasmid-encoded *virB* and *trb* operons. The *virB* genes and some of the known functions of the encoded products are presented at the top. This T4SS is closely related in operon organization and subunit composition to a T4SS encoded by the *E. coli* conjugative plasmid pKM101. The Trb system is closely related in operon organization and subunit composition to a T4SS encoded by the *E. coli* conjugative plasmid pKM101. The Trb system is closely related in operon organization and subunit composition to a T4SS encoded by the *E. coli* conjugative plasmid pKM101. The Trb system is closely related in operon organization and subunit composition to a T4SS encoded by the *E. coli* conjugative plasmid RP4. Genes encoding protein homologs are identically color-coded.





Figure 4 | **Structure of a type IV secretion core complex.** The core complex<sup>53</sup> is composed of TraN (a VirB7 homologue), TraO (a VirB9 homologue) and TraF (a VirB10 homologue), which are encoded by the *Escherichia coli* conjugative plasmid pKM101. This structure was obtained using cryo-electron microscopy and is viewed from the side (upper left panel), tilted towards the outer membrane side (lower left panel) and tilted towards the inner membrane side (lower right panel). The cut-out view (upper right panel) details the proposed transmembrane regions and the localization of the VirB7, VirB9 and VirB10 homologues within the structure. C, carboxy-terminal domain; N, amino-terminal domain.



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Agrobacterium transformation of plant cells is mediated by intercellular signalling, attachment, virulence protein catalysed DNA transfer to the nucleus and genome integration.

Removal of the tumour-forming genes, and separation of the virulence functions (Vir genes) on a separate "helper" plasmid allows simpler manipulation of the T-DNA and genes to be inserted into the plant genome.





### "Disarmed" binary plasmids



#### Summary of Agrobacterium mediated gene transfer and plant regeneration







A







D



Е



F

Figure 4 Regeneration of transgenic maize plants







Figure 3. Important Historical Milestones in Plant Transformation.

Since its beginning in 1977, the pace of crop transformation technology development has not been linear. In recent years, the genome editing revolution begs for crop transformation improvements to enable greater food security.